Bursting properties of units in cat globus pallidus and entopeduncular nucleus: the effect of excitotoxic striatal lesions

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The bursting properties of units recorded in globus pallidus and entopeduncular nucleus were studied in awake cats sitting quietly before and after ipsilateral excitotoxic striatal lesions. A computerized statistical procedure was used to identify and evaluate bursts in the recorded spike trains. Bursts were assigned a quantitative statistical measure of burst 'strength' (or improbability) – the surprise value. Before the lesion, 34% of units in the globus pallidus and 60% of units in the entopeduncular nucleus exhibited bursts. Burst units had a significantly slower discharge rate and a significantly greater variability of discharge than non-burst units. The mean length of the interspike intervals immediately preceding the bursts was significantly longer than the overall median intervals in burst units. After the lesion, 21% of units in the globus pallidus and 11% of the units in the entopeduncular nucleus exhibited bursts. Burst units had significantly higher discharge rates and lower discharge variability after the lesion. In contrast, the lesion had no significant effect on the rate or variability of non-burst units. The differences between bursting and non-bursting units in discharge rate and variability disappeared after the lesion. In globus pallidus, the lesion resulted in a significant reduction in the mean number of bursts per unit, surprise value per burst, mean length of bursts, and number of spikes per burst, and a significant increase in the mean discharge rate of burst units. In entopeduncular nucleus, the small number of bursts recorded after the lesion precluded a useful statistical comparison of the effect of striatal lesions on the properties of the bursts. This study demonstrates that removing striatal projections to globus pallidus and entopeduncular nucleus decreases bursting in these nuclei, indicating that intact striatal projections are necessary for the normal production of bursts in these regions.

INTRODUCTION

Neuronal burst discharges are thought to be important in transmitting information in the nervous system and have been implicated in sleep, epilepsy, and long-term potentiation. Bursting has been found in studies of single unit discharge in the globus pallidus (GP) and entopeduncular nucleus (EPN) of the cat. Neurons in the striatum utilizing γ-aminobutyric acid (GABA) as a neurotransmitter project to GP and EPN, and evidence suggests that the neurotransmitter GABA may be involved in neuronal bursting. Striatal stimulation or application of GABA to pallidal neurons inhibits GP neuronal discharge. In addition, application of GABA on thalamic, substantia nigra and hippocampal neurons reduces neuronal excitability and bursting. The application of GABA antagonists on hippocampal neurons increases burst discharges in these neurons. Striatal lesions produced with the excitotoxin ibotenic acid result in a 90–95% decrease in extracellular GABA in striatal targets. Thus, it might be expected that striatal lesions would increase bursting in GP and EPN. Excitotoxins, which destroy all classes of striatal neurons and leave the fibers of passage intact, are thought to produce neuropathological and neurochemical changes similar to those seen in Huntington's disease. Thus, issues related to Huntington's disease can be addressed in studies of animals with excitotoxic striatal lesions.

In the present study we have examined the effects of excitotoxic lesions of the striatum on burst discharges in the GP and EPN. The present study presents additional analysis of data presented in a recently published report. In this report, the effects of striatal lesions on tonic discharge properties of neurons in the pallidum were analyzed. This work indicated that the variability of interspike intervals in the GP and EPN decreased after striatal lesions. Alterations in the variability of discharge might be related to altered bursting properties of neurons in the pallidum. Thus, in the present study we examined the influence of striatal lesions on neuronal burst discharges in GP and EPN.

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MATERIALS AND METHODS

Procedures

Four cats were habituated to the recording procedures and trained to sit quietly in a loose restraint. Two to three weeks later, they were prepared surgically for chronic unit recordings. In each animal, a chamber was positioned stereotaxically and fixed to the left side of the cranium. The chamber provided access to the GP, EPN, caudate nucleus and putamen. One week after surgery, the animals were reh abitudinated to the recording apparatus and exploratory recordings were initiated to locate the pallidum and striatum. Data collection with tungsten microelectrodes began 2–3 weeks after surgery. Computerized procedures for digitizing spikes on-line assisted with the recording[3]. Units recorded at each sector were discriminated from each other and from background noise on the basis of their negative peak amplitudes, positive peak amplitudes, and peak-to-peak times[39]. Discriminated units were used in subsequent analyses. The results presented in this paper represent additional analysis of data presented in a recently published paper[39].

Spontaneous unit activity was recorded from the GP and EPN of alert cats sitting quietly. The animals were loosely restrained with a plastic collar. The behavioral state of the cats was monitored by EEGs, movement detectors (accelerometers or high-impedance cables), video monitors, or a combination of these. Unit activity occurring during movement or 500–1500 ms before or after a movement was discarded. In the first two cats, EEG activity was recorded to identify slow waves, large amplitude signals indicating early sleep stages. Since the cats were usually awake during the short (less than 2 h) recording sessions and since visual observations proved as useful as EEGs, the subsequent two cats were monitored with a video camera alone. The first two animals were studied both before and after the lesion. In these cats, penetrations through GP and EPN were made in a checkerboard fashion. A set of recording tracks through the GP and EPN was studied before the lesion. The remaining tracks neighboring those studied before the lesion were examined after the lesion. This procedure of recording from neighboring tracks before and after the lesion ensures that unit data are gathered from similar sites. One of the remaining two cats was studied only before the lesion, and the other was studied only after the lesion. This procedure of studying one cat only after the lesion provided some control for the effects of electrode damage on the discharge in GP and EPN. Damage caused by the electrode had no noticeable effect on the discharge pattern in GP and EPN, as pre-lesion and post-lesion data obtained from animals studied both before and after the lesion were similar to that obtained from animals studied only before or after the lesion. Unilateral striatal lesions were produced by multiple injections of ibotenic acid (2 μg/200 μl) into the caudate nucleus and putamen. Eleven to 19 injection sites ranging from 0.25 to 1.5 μl/site were used to produce the lesions. Cell loss in the striatum is thought to occur within a week after injection of the excitotoxin[3]. Thus, in the present study unit activity in the pallidum was recorded between 7 and 100 days after the lesion.

Histologic reconstructions of the recording sites were facilitated by marking 6 recording tracks with deposits of iron on the final day of recording. Direct current (100 μA for 30–45 s) was passed through a bipolar steel electrode, providing iron deposits that were stained blue by adding 1.5% potassium ferrocyanide to the 10% formalin perfusing solution. The animals were perfused transcardially under deep barbiturate anesthesia. The brains were removed, postfixed in sucrose/formalin solution, frozen, sectioned, and stained with Cresyl violet. The recording tracks and lesions were reconstructed and drawn on maps traced from stained sections. The lesion size was determined with the aid of an image analysis computer. An outline of the striatum was traced on a digitized image of representative coronal sections. The striatal areas on the side of the lesion and on the contralateral side were computed and compared.

Data analysis

Conventional techniques such as the autocorrelation method or examination of interval distributions cannot detect individual bursts in a spiketrain. Consequently, procedures similar to those described by Legendy and Salzman were used to detect bursts[35]. A group of spikes (n > 3) was initially identified as a burst if the mean rate of activity for the group of spikes was at least twice that of the mean rate for the whole spiketrain. Bursts were assigned a quantitative, statistical measure of burst strength – the Poisson surprise value – defined as the negative logarithm of the probability of the occurrence of a burst in a random (Poisson) spiketrain. Following the initial calculation of the surprise value, this value was maximized by adding an additional spike to the end of the burst to determine whether this would increase the surprise value. If adding the spike increased the surprise value, the spike was included in the burst. This process of adding spikes if they increased the surprise value was repeated as long as the surprise value increased. Maximization of the surprise value was pursued further by removing spikes from the beginning of the burst. If removing a spike increased the surprise value, the spike was removed from the burst. The surprise value for each burst, the length of each burst, the number of spikes in each burst, the rate within each burst, and the number of bursts per 1000 spikes (burst rate) were calculated. A group of spikes was considered to be a burst if the Poisson surprise value was greater than 3, indicating that the particular group of spikes had a less than 1 in 1000 chance of being a random event. To determine the length of the interval preceding the burst, the burst and the interval preceding the burst were displayed graphically. The intervals were then directly measured from the print-out. For long pre-burst intervals (400–2000 ms), the actual interval could exceed the displayed value. Nevertheless, in these cases the displayed value was used as an estimate of the interval. This method tends to underestimate the length of the long pre-burst intervals. The median and coefficient of variation (standard deviation/mean) of the interspike intervals for all units were also calculated.

The Mann–Whitney U-test and the chi-square test were used to assess the significance of changes in bursting. The binomial test was

![Fig. 1. Representation of recording sites where bursts were detected. Pre-lesion recording sites from all animals are plotted as filled circles and post-lesion recording sites from all animals are plotted as dashes. Units exhibiting no bursts were interspersed with the units exhibiting bursts.](image-url)
used to assess the significance of the difference between the median interval of the unit and the length of the interval immediately preceding the burst. To determine whether the surprise value was linearly correlated to any of the other properties of a burst, correlation coefficients were also calculated.

**TABLE 1**

Discharge properties of units in globus pallidus

<table>
<thead>
<tr>
<th></th>
<th>Pre-lesion</th>
<th>Post-lesion</th>
</tr>
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<tbody>
<tr>
<td>Bursting</td>
<td>Non-burst</td>
<td>Bursting</td>
</tr>
<tr>
<td>Median interval (ms)</td>
<td>37 (3.3)</td>
<td>35 (2.1)</td>
</tr>
<tr>
<td>Mean interval (ms)</td>
<td>94 (12)</td>
<td>45 (4.6)*</td>
</tr>
<tr>
<td>Coefficient of variation</td>
<td>2.1 (0.2)</td>
<td>0.9 (0.02)*</td>
</tr>
<tr>
<td>Number of units</td>
<td>66</td>
<td>126</td>
</tr>
</tbody>
</table>

* P < 0.001 as assessed by the Mann–Whitney U-test. ms = milliseconds.

**RESULTS**

**Striatal lesion**

The striatal lesion extended through the entire rostro-caudal extent of the caudate nucleus in all animals. The rostral putamen was also lesioned in all cats. In each section examined, the lesion resulted in a 25–50% loss of striatal tissue on the lesioned side as compared to the contralateral side. The lesion produced no noticeable

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**Fig. 3.** Distribution of median interspike intervals in globus pallidus before (A) and after (B) striatal lesion. Distribution of coefficients of variation of the interspike intervals before (C) and after (D) striatal lesion. The distributions of the median interspike intervals and coefficients of variation include only units exhibiting bursts.
behavioral, motor, or postural changes in any animal.

Site of recording

Burst units in the GP and EPN (Fig. 1) were detected in similar regions before and after the lesion. Burst units were intermingled with non-burst units.

Globus pallidus

Prior to the lesion, one third of the units in GP (66 of 192) exhibited bursts (Fig. 2). On average, both burst units and non-burst units had similar median interspike intervals, however, the mean interspike intervals of burst units were significantly longer than those of non-burst units (Table I). Burst units had significantly greater variability of discharge than non-burst units, i.e. burst units had larger coefficients of variation than non-burst units (Table I).

The striatal lesion had no effect on the overall discharge rate or variability of non-burst units (Table I), but the lesion had substantial effects on burst units. The lesion resulted in a 14% decrease in the proportion of units exhibiting bursts ($P < 0.01$, chi-square test); after the lesion only 55 of 265 units exhibited bursts (Fig. 2). Prior to the lesion, burst units had significantly longer median intervals and significantly greater variability than burst units after the lesion ($P < 0.0001$, Mann–Whitney U-test). The lesion resulted in a 32% decrease in the length of the median interval and a 33% decrease in the coefficient of variation (Table I). Examination of the distribution of median intervals and coefficients of variation suggests that the proportion of burst units with long median intervals (Fig. 3A,B) or large coefficients of variation (Fig. 3C,D) decreased considerably. Concomitant to the decreased variability, the rate of bursting (bursts/1000 spikes) in neurons exhibiting bursts also decreased by 23% after the lesion (Table II).

The striatal lesion altered the characteristics of the bursts (Fig. 4). The mean burst length was significantly reduced (Table II), as demonstrated by a decreased proportion of bursts greater than 80 ms in duration (Fig. 4A,B). The mean number of spikes per burst was

<table>
<thead>
<tr>
<th>TABLE II</th>
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Properties of bursts in globus pallidus and entopeduncular nucleus

Numbers in brackets represent standard errors.

<table>
<thead>
<tr>
<th></th>
<th>Globus pallidus</th>
<th>Entopeduncular nucleus</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Lesion</td>
</tr>
<tr>
<td>No. of spikes</td>
<td>14 (1.9)</td>
<td>8 (0.4)*</td>
</tr>
<tr>
<td>Intra-burst rate (hz)*</td>
<td>183 (35)</td>
<td>251 (23)*</td>
</tr>
<tr>
<td>Burst duration (ms)</td>
<td>200 (30.5)</td>
<td>43 (5.6)*</td>
</tr>
<tr>
<td>Surprise value</td>
<td>8 (0.8)</td>
<td>4 (0.13)*</td>
</tr>
<tr>
<td>Bursts/1000 spikes</td>
<td>14 (2.2)</td>
<td>11 (2.6)</td>
</tr>
<tr>
<td>No. of bursts</td>
<td>273</td>
<td>158</td>
</tr>
<tr>
<td>No. of units</td>
<td>55</td>
<td>65</td>
</tr>
</tbody>
</table>

* Computing an intra-burst rate from the mean values of the number of spikes and burst duration gives different results for intra-burst rate than the mean computed from individual bursts (shown). The former reduces the impact of the short bursts with few spikes, especially when these bursts have a high intra-burst rate.

* $P < 0.0001$ as assessed by the Mann–Whitney U-test.

ms = milliseconds, hz = spikes/s.

Fig. 4. Examples of bursts in GP and EPN. Pre-lesion GP (A–D), pre-lesion EPN (E and F) and post-lesion GP (G and H). Arrowheads mark the beginning and end of bursts. Bursts recorded after the lesion are shorter and contain fewer spikes than bursts recorded before the lesion. In trace F, there are two bursts.
reduced by 43% (Table II). The distributions of the number of spikes in bursts demonstrate that, after the lesion, the proportion of bursts containing many spikes (n > 10) decreased (Fig. 3C,D). The lesion also resulted in a significant increase in the rate of discharge within bursts (Table II), and the distributions of the rates suggest that the proportion of bursts with a high discharge rate increased after the lesion (Fig. 5E,F). The average surprise value, an indication of burst strength (or burst improbability), decreased significantly after the lesion (Table II). This was most evident from the decreased proportion of bursts with large surprise values (Fig. 6A,B). The decreased surprise value was probably related to the decreased number of spikes per burst since the correlation between the surprise value and the number of spikes in the burst was so strong (correlation coefficient, r = 0.95). The surprise value was not as well correlated to the length of the burst (r = -0.4), the rate of discharge in the burst (r = -0.1), the mean interval length (r = 0.1), or the median interval length (r = -0.1) of the units (Fig. 7). The intra-burst rate was weakly related to the overall discharge rate (r = 0.5).

GP bursts were frequently preceded by long intervals (Fig. 4). These intervals were significantly longer than the median interval for the unit (P < 0.01, binomial test). The mean pre-burst interval was 99 ms (S.E.M. = 16), about 3 times longer than the average median interval. Following the lesion the pre-burst interval decreased significantly to 78 ms (S.E.M. = 8.9) (P < 0.005, Mann–Whitney U-test). Even after the lesion, however, the pre-burst intervals were significantly (3 times) longer than the median interval (P < 0.01, binomial test).

![Images of graphs and figures](Figures)

Fig. 5. Distribution of burst duration, number of spikes, and surprise values in globus pallidus bursts before and after the striatal lesion. Burst duration pre-lesion (A) and post-lesion (B). Number of spikes in the burst pre-lesion (C) and post-lesion (D). Intra-burst rate pre-lesion (E) and post-lesion (F). Note that the post-lesion distributions of burst duration and number of spikes shift towards shorter values whereas the post-lesion distribution of rates shifts towards larger values.
**Entopeduncular nucleus**

Prior to the lesion, a majority (60%) of units exhibited bursts (Fig. 2). On average, the interspike intervals of burst units were significantly more variable ($P < 0.001$, Mann–Whitney $U$-test) than non-burst units (Table III). Burst units also had significantly lower mean rates of discharge ($P < 0.001$, Mann–Whitney $U$-test) than non-burst units (Table III). Following the striatal lesion the proportion of units exhibiting bursts decreased markedly ($P < 0.01$, chi-square test), to only 11% (Fig. 2). The small number of bursts recorded after the lesion precluded a useful statistical comparison of the effect of the striatal lesion on the properties of bursts in EPN. As in GP, however, the lesion did not affect the overall discharge properties of non-burst units (Table III).

**Prelesion comparison of globus pallidus and entopeduncular nucleus**

Prior to the lesion, a larger proportion of burst units in EPN had small median intervals ($P < 0.001$, Mann–

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Fig. 6. Distribution of surprise values in globus pallidus pre-lesion (A) and post-lesion (B).

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Fig. 7. Scatter plots of pre-lesion globus pallidus burst parameters: A: number of spikes in burst vs. surprise; B: intra-burst rate vs. surprise; C: mean interval vs. surprise; and D: intra-burst rate vs. overall mean rate. The values illustrated in the scatter plots are culled from 175 globus pallidus bursts. The surprise values are linearly related to the number of spikes in the burst.
Whitney U-test) as compared to GP (compare distributions in Fig. 8A and Fig. 3A). Burst units in EPN were also less variable ($P < 0.0001$, Mann–Whitney U-test) than burst units in GP (Fig. 8B vs. Fig. 3B). Bursts in EPN had significantly smaller mean surprise values ($P < 0.005$, Mann–Whitney U-test) than bursts in GP (Table I). Bursts with small surprise values (surprise < 5) were more common in EPN than in GP (Fig. 8C vs. Fig. 5E).

**DISCUSSION**

In this study, we found that excitotoxic lesions of the striatum resulted in decreased neuronal bursting in GP and EPN. In GP, the lesions resulted in a decreased proportion of units exhibiting bursts and in a higher overall discharge rate and lower variability in burst units.
In EPN, the striatal lesions resulted in a dramatic decrease in the proportion of units that burst. The lesions had no effect on the properties of non-burst units in either GP or EPN.

In GP, the reduction in bursting following the striatal lesion was accompanied by an increased intra-burst discharge rate, decreased burst length, fewer spikes per burst and decreased surprise values of the bursts. A decreased overall mean interval length (i.e. an increase in the discharge rate) in burst units and an increased rate within the burst are both consistent with the loss of a tonic inhibitory input from the striatum. The decreased variability in post-lesion spike trains is probably related to a reduced number of bursts. The decreased surprise value, decreased length of bursts, and decreased number of spikes per burst all suggest that GP bursts are ‘weaker’ after the lesion. These results indicate that striatal inputs are important for production of ‘normal’ bursts in GP and EPN. It may be that partially deafferented GP and EPN neurons are unable to produce ‘normal’ bursts. Alternatively, these changes in bursting may reflect a compensatory response to the lesion.

The increased post-lesion mean discharge rate in GP could potentially affect the detection of bursts in GP. The detection of bursts with the surprise technique depended on the mean discharge rate of individual spike trains. Initially, a group of 3 spikes in a spike train was considered to be a burst if that group had a mean discharge rate that was at least twice the mean discharge rate for the spike train. Thus, the increased post-lesion discharge rate in GP might by itself reduce the number of bursts detected in GP after the lesion. To be classified as a burst, however, that group of 3 spikes also had to have a surprise value of at least 3, which usually meant an addition of spikes. The number of spikes per burst usually exceeded 3. Thus, the direct relationship between the mean discharge rate and the properties of bursts was weakened. Data from EPN indicate that the reduction in the number of bursts detected in EPN occurs independently of any changes in the mean discharge rate. The number of bursts detected in EPN decreased after the lesion even though the mean discharge rate was not affected by the lesion. Moreover, the huge difference between the mean discharge rate and the mean intraburst rate in GP suggests that post-lesion alterations in the mean discharge rate have little effect on the detection of bursts. The pre-lesion intraburst rate in GP is 17 times the mean discharge rate in GP. This difference in the pre-lesion intra-burst rate and the mean rate is much larger than the difference between the pre-lesion and post-lesion mean rates.

Legendy and Salcman developed the ‘Poisson surprise’ procedure for detecting and evaluating bursts in the visual cortex of alert cats. They used a surprise value of 10 as a cut-off for selecting bursts. Bursting in cat GP and EPN is different from bursting in the visual cortex. Spike trains in the visual cortex exhibit more bursts and have slower rates of discharge than those in the pallidum. The burst discharges in visual cortex have on average larger surprise values than bursts in GP and EPN. The mean discharge rate in visual cortex bursts is on average only 3–6 times higher than the overall mean discharge rate whereas the mean discharge rate in GP and EPN bursts is on average 7–10 times the overall median discharge rate. The surprise cut-off in the present study was 3, which might result in the inclusion of less intense bursts in our sample as compared with the sample from visual cortex. It is possible that the differences in bursting reflect differences in inputs, receptors, and transmitters found in these two regions. For example, the cat pallidum probably receives only one major glutamatergic input and has few glutamate receptors, whereas the visual cortex receives many glutamatergic inputs and has a large number of glutamate receptors. Glutamate is an excitatory transmitter thought to produce burst firing.

In the present study, we excluded data associated with limb and body movements because these movements are thought to alter the pattern of unit activity in the basal ganglia. Our goal was to study spontaneous discharge with a constant behavioral baseline. To this end, we trained the animals to sit quietly, and monitored their movements to exclude any data collected during movements. In contrast, the goal in the work of Legendy and Salcman was to develop a technique for evaluating bursts, therefore, they did not restrict visual stimulation or eye movements, which are thought to trigger burst discharges in the visual cortex. In fact, in Legendy and Salcman’s study, visual stimulation elicited most of the burst discharges. This fundamental difference in the recording paradigm and goals of the two studies may be yet another reason for the observed differences in bursting.

Previous studies of unit activity in the cat pallidum identified burst discharges in both GP and EPN. Previous investigations in monkey and the present study in cat demonstrate that a significant proportion of units in the lateral GP of monkey and in the GP of cat exhibit bursts and burst units in both cat and monkey have much slower mean rates of discharge than non-burst units. No bursts have been described in units of the medial GP of monkey. In the present study, however, we find that the EPN has burst units. Besides species differences, methodologic differences may account for the results. Earlier studies did not have the advantages of the surprise technique, which provides detailed information
about the characteristics of individual bursts. In the earlier studies, bursting was assessed by examining interval distribution characteristics, i.e. mode, and the mean rate of discharge. Interval distribution characteristics and even autocorrelation measures provide no information about individual bursts in a spiketrain.

Our work suggests that units in GP have different bursting characteristics than those in EPN. GP bursts are more prominent than EPN bursts, i.e. they are longer, contain more spikes, and have larger surprise values. Consequently, bursts in GP are more noticeable than bursts in EPN, and this may account for the absence of bursting in the earlier descriptions of unit discharge in the monkey medial globus pallidus. The different peptidergic inputs from striatum to GP and EPN may explain the differences between the bursts that normally occur in GP and EPN and may also explain why bursting in EPN is reduced compared to GP. Enkephalin, the peptidergic transmitter co-localized with GABA in striatal inputs to GP, is thought to be inhibitory whereas substance P, the peptidergic transmitter found in the striatal inputs to the EPN, is thought to produce long slow depolarizations conducive for bursting. The larger proportion of bursting units detected in EPN as compared to GP suggests that substance P may have a role in burst production.

Decreased bursting in GP and EPN after the striatal lesion is not consistent with our initial prediction, which was based on the findings that (1) GABA application reduces neural excitability and bursting and (2) decreased extracellular GABA in GP and EPN results from excitoexcitotoxic striatal lesions. On the contrary, our results suggest that striatal input is necessary for the normal production of bursts in GP and EPN. The idea that GABA is responsible for bursts in these structures is supported by the observation that the pre-burst interval in GP and EPN is usually longer than the median interval for the unit, suggesting that GABAergic inhibition results in the long interval preceding a burst. The long inhibition may then be followed by rebound bursting in GP and EPN, similar to that seen in thalamus. Even though the GABA-mediated mechanism of rebound bursting is consistent with our results, other GABAergic mechanisms may be important in bursting. It may be that, as in the thalamus, the resting membrane potential of a neuron determines whether a neuron bursts. The more hyperpolarized the resting membrane potential of a thalamic neuron, the more likely it is to burst when depolarized. A similar mechanism may work in GP and EPN. Perhaps, following the striatal lesion, the decrease of GABA in GP and EPN results in a chronically more depolarized resting membrane potential, thereby reducing bursting.

Another mechanism may also be important in explaining decreased bursting in GP and EPN after striatal lesion. Recent work in the hippocampus suggests that a reduction in inhibition by itself is not sufficient for increased bursting. In addition to reduced inhibition, excitatory inputs must be active for bursting to occur. In our study, it is likely that the excitatory inputs to GP and EPN were affected by the striatal lesion. The majority of excitatory inputs to GP and EPN are from the subthalamic nucleus and this nucleus in turn receives inhibitory inputs from GP. Our work demonstrates that striatal lesions increase the discharge rate in GP. This increase presumably enhances inhibitory input to the subthalamic nucleus, probably causing subthalamic neurons to discharge more slowly. This would reduce the number of EPSPs impinging on GP and EPN neurons, and would reduce bursting in these nuclei even though the amount of GABAergic inhibition is decreased in both nuclei.

Hypotheses attempting to explain the mechanisms underlying the chorea or bradykinesia of Huntington’s disease have focused on the effects of striatal lesions on discharge rates in pallidum, VA/VL thalamus and cortex. Our results suggest that changes in burst discharges may also be important.

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